

web: <https://bonn.leibniz-lib.de/en/biobank>

email: biobank@leibniz-lib.de

phone: +49 (0)228 9122-357, -355

address: Museum Koenig, Adenauerallee 127, 53113 Bonn, Germany | visitors: Adenauerallee 160

LIB Biobank: deposition of tissue / DNA / cells

You are considering material donations to the LIB Biobank – thank you for your interest and trust!



- The LIB Biobank stores samples derived from **animals** (no human, bacterial etc. samples) and **environmental samples** (e.g. eDNA, community DNA, soil samples, gut samples).
- We accept samples that have been preserved for molecular analysis (e.g. flash-frozen **tissue**; tissue in 96% ethanol, DMSO, RNAlater; extracted **DNA**, RNA, etc.; also **viable cells**).
- Samples should be **documented extensively** (taxonomic determination ideally down to species or genus level; detailed collecting/field information; **permits**, if necessary → Nagoya-relevant, CITES, collecting, etc.) and must be free from third-party claims.
- Except for eDNA etc., the samples should also be referenced to a **voucher specimen** (for morphological examination) deposited in a public collection (e.g., LIB museums in Bonn or Hamburg), or can alternatively be submitted as whole organisms if small enough (see below). If there is no other way, high-quality, informative photographs can function as e-vouchers instead of a physical voucher.
- Samples can be blocked for requests from third parties for a specified period. Should they wish so, sample donors can also be asked for their consent when a request for the respective sample is made.

The following page contains suggestions / details on animal tissue preservation and submission.

- Pre-labeled, cryogenic 2 mL **collecting tubes can be provided** by us. We also have an alternative format (for limited sample numbers) with 5 mL tubes.
- Samples should be **fresh** or only a **few years old**. In case of doubt, we can test DNA integrity.
- Water/humidity and elevated temperature are the greatest threats to DNA preservation:
 - **Short-term storage:** keep samples **in refrigerator** (do not freeze).
 - **Medium-term storage**, from ~3 weeks on: preferably **freeze and do not thaw** until delivery; **repeated freezing/thawing damages** the DNA considerably.
- Samples for **DNA analysis** should be stored **in 96% ethanol as soon as possible**.
 - **The ratio of sample:ethanol should not exceed ca. 1:10** (high 'excess' of ethanol necessary); otherwise, please replace ethanol after a few hours so it is not diluted.
 - **Avoid low percentage alcohol!** The water in it leads to DNA degradation; 100% ethanol may contain traces of desiccants: 96% ethanol is optimal.
 - If at all possible, use **non-denatured alcohol only**.
 - Avoid contact with formalin (formol) and solutions of acetic acid (or any other acid).
- Samples for **RNA analysis** should be stored in **RNAlater** (if large, partly homogenized) and frozen as soon as possible (after 1 night); for 'homebrew' RNAlater, please provide the recipe
- Other preservation fluids may be suitable as well (e.g. propylene glycol), please contact us.
- If you would like to supply **viable cell samples** (e.g. from animals not dead more than ~2 days), please contact us beforehand: cells@leibniz-lib.de and **do not freeze**.
- For LIB staff: we can provide **dry/vapor shippers** for transport (also by plane) of samples at liquid nitrogen temperatures. These can also be used to directly snap freeze the samples in the field, e.g. **in case you want to avoid exposure to preservation fluids** by maintaining an uninterrupted cold chain (shippers hold the temperature for ~1.5 weeks).
- Small animals (up to ca. 4 cm) can enter the biobank as **complete individuals**. Cuts/punctures into the tissue or removal of a leg/s enable quicker penetration by the fluid.
- If you wish to split the material, or if you have larger animals, please collect them as for morphological examination and immediately preserve molecular **subsamples, e.g. arthropod legs** (2-3 from ideally the right side of the body) **or muscle** tissue into tubes with preservation fluid. Sterilize instruments after subsampling (please ask if you do not know how).
- **Do not put more than one animal into a single tube**; never mix subsamples from 2 animals.
- If you have to use a killing jar, let the killing medium act aerielly (e.g., a droplet on cotton) and - very importantly - do not leave the sample in the jar for more than max. very few hours (enzymes swiftly eat away at the DNA). Always **avoid direct sun** exposition or hot vehicles.
- Vertebrate tissue samples:
 - **muscle tissue** preferred. Avoid slimy/mucous or hardened tissue and esp. intestines.
 - Preserve a **tissue piece of max. 0,5cm³**, ideally as several small pieces; additional tissue from the same animal can go into a second or third tube.
 - Easy-to-sample tissues: amphibians/reptiles/birds: toes, toepads, tip of tail (if any); small mammals: piece of ear or tail; fish: eye, pectoral fin.
 - For **cell culture**: as above, but preferably **skin and eyes**. **Sterilize thoroughly** (with 70% ethanol). We provide you with cell media for sampling/shipping. Do NOT freeze.
- We will ask you for the metadata of your samples (taxonomy, collecting data); you can submit your own tables or better: we provide our scaffold in which you see all available data categories (using this minimizes questions from our side).
- We gladly accept photographs of specimens (or habitats) and can give you the specifications.

